

**Biosynthesis of the Shikimate-derived Starter Unit  
of the Immunosuppressant Ascomycin:  
Stereochemistry of the 1,4-Conjugate Elimination**

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(Received for publication April 14, 1997)

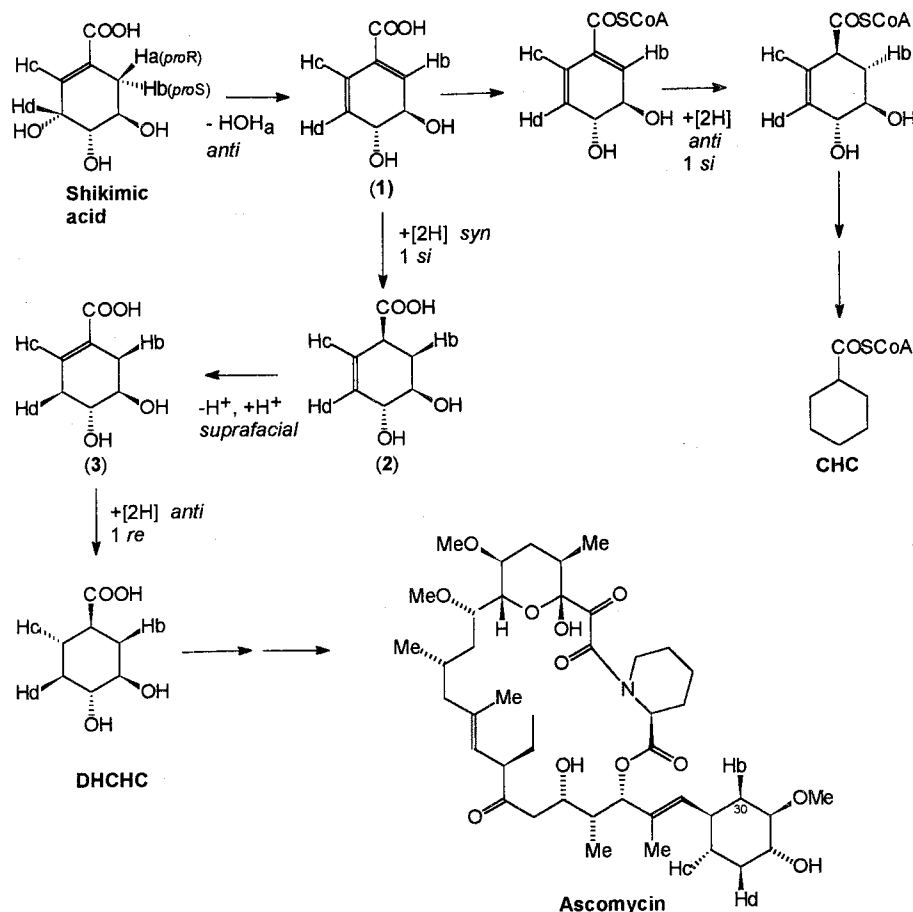
The potent immunosuppressants ascomycin, FK506 and rapamycin are polyketide secondary metabolites that are biosynthesized from an usual shikimate-derived (1*R*,3*R*,4*R*)-3,4,-dihydroxycyclohexanecarboxylic acid (DHCHC) starter unit<sup>1-4</sup>). The biosynthetic pathway to this DHCHC moiety of ascomycin has been investigated in *Streptomyces hygroscopicus* var. *ascomyceticus* by a series of incorporation experiments with isotopically labeled shikimic acid and various putative pathway intermediates<sup>1</sup>). The results obtained were consistent with a process that initiates with either a *syn* or an *anti* 1,4-conjugate elimination of a C6 hydrogen and the C3 hydroxyl group of shikimic acid to produce (3*R*,4*R*)-3,4-dihydroxy-1,5-cyclohexadienecarboxylic acid (**1**) (Fig. 1). A *syn* reduction of the  $\Delta^1$ -double bond of **1** gives of (1*S*,4*R*,5*R*)-4,5-dihydroxycyclohex-2-enecarboxylic acid (**2**), which is subsequently converted to (4*R*,5*R*)-4,5-dihydroxycyclohex-1-enecarboxylic acid (**3**) by an isomerization of the remaining double bond from the  $\Delta^2$  to the  $\Delta^1$  position. In the final step of the process **3** is converted to DHCHC by an *anti* addition of hydrogen to the *re* faces of C1 and C2. An alternative pathway in which the first double bond is reduced by an *anti* addition of hydrogen (**1**→**2**), while consistent with the experimental results, seems unlikely as it would require a subsequent unprecedented antarafacial rearrangement to generate **3**<sup>1</sup>).

The only stereochemical detail of DHCHC biosynthesis that remains undetermined is the steric course of the initial 1,4-conjugate elimination. Previous incorporation studies with (–)-6(*R,S*)-[6-<sup>2</sup>H<sub>1</sub>]shikimic acid had generated ascomycin which exhibited a single <sup>2</sup>H single at 0.9 ppm, consistent with deuterium occupy-

ing the C3*R* (axial) position<sup>1</sup>). This signal clearly demonstrated that the pathway to DHCHC involves loss of one hydrogen (no signal was observed for deuterium occupying the C3*S* position), but it does not distinguish which of the heterotropic C6 hydrogens is removed. We have now circumvented this problem by developing a synthetic strategy for preparing stereospecifically deuterated (–)-6(*R*)-[6-<sup>2</sup>H<sub>1</sub>] and (–)-6(*S*)-[6-<sup>2</sup>H<sub>1</sub>]shikimic acid from mannose<sup>5</sup>). In separate experiments each stereospecifically labeled shikimic acid (30 mg) was added to 5 × 25 ml 24 hours cultures of *S. hygroscopicus* var. *ascomyceticus* to a final concentration of 1.4 mM. After an additional 5 days of fermentation the cells were combined with those obtained from a 2 × 25 ml six day fermentation, and the ascomycin was isolated, purified and characterized by <sup>1</sup>H and <sup>2</sup>H NMR as described previously<sup>1</sup>). Inspection of the 46.07 MHz <sup>2</sup>H NMR spectra of ascomycin (71 mg) from the feeding experiment with (–)-6(*R*)-[6-<sup>2</sup>H<sub>1</sub>]shikimic acid exhibited no discernible signal even after prolonged accumulation. In contrast, the <sup>2</sup>H NMR spectrum of ascomycin (56 mg) from the (–)-6(*S*)-[6-<sup>2</sup>H<sub>1</sub>]shikimic acid experiment was indistinguishable from that obtained previously using (–)-6(*R,S*)-[6-<sup>2</sup>H<sub>1</sub>]shikimic acid, with a single <sup>2</sup>H signal at 0.9 ppm.

These results clearly indicate that the initial 1,4-elimination step proceeds in an *anti* fashion with loss of the *pro*-6*R* hydrogen of shikimate. The same *anti* elimination has been shown to occur in the conversion of shikimate to **1** during the biosynthesis of cyclohexanecarboxylic acid (CHC), a process which occurs in *Streptomyces collinus* and *Alicyclobacillus acidocaldarius* (Fig. 1)<sup>5</sup>). The CHC and DHCHC pathways apparently diverge after this first step. Analysis of the rapamycin polyketide synthase provides strong evidence that this enzyme utilizes DHCHC as a free acid suggesting that the *syn* reduction of the  $\Delta^1$ -double bond of **1** in this pathway occurs with the free acid<sup>3,6</sup>). Evidence indicates that in the CHC pathway **1** is first converted to a coenzyme A thioester and that the  $\Delta^1$ -double bond of this compound is then reduced in an *anti* fashion (Fig. 1)<sup>7-9</sup>). It has been proposed that these stereochemical differences reflect differing progenitors for the enoyl reductases in these two pathways<sup>10</sup>). By the same token the dehydratases responsible for converting shikimate to **1** in the same stereochemical fashion in these two pathways may be related. It has previously been noted that chorismate synthase also catalyzes an *anti* 1,4-elimination reaction using 5-enolpyruvylshikimate

Fig. 1. Proposed commonality in the initial step of the pathways to DHCHC and CHC from shikimic acid.



3-phosphate in a later step in the shikimate pathway<sup>5</sup>). The evolutionary relationship of all of these dehydratases, however, will only be determined once the corresponding genes have been cloned. Some progress has been made in this direction; an enoyl CoA reductase involved in the CHC pathway has already been cloned and sequenced<sup>7</sup>), and analysis of the surrounding genes has provided evidence of a CHC biosynthetic gene cluster (unpublished data).

#### Acknowledgment

This work was supported by grants from the National Science Foundation (MCB-9418581 to KAR) and the National Institutes of Health (AI202640 to HGF). We thank Dr. STEFAN GRÖGER for recording the NMR spectra.

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